



## Co-opting Moore's law: Therapeutics, vaccines and interfacially active particles manufactured via PRINT®



Joseph M. DeSimone

Division of Molecular Pharmaceutics, Eshelman School of Pharmacy, Lineberger Comprehensive Cancer Center, Department of Chemistry, University of North Carolina, Chapel Hill, NC 27599, USA  
 Department of Chemical and Biomolecular Engineering, NC State University, Raleigh, NC 27695, USA  
 Sloan-Kettering Institute for Cancer Research, Memorial Sloan-Kettering Cancer Center, New York, NY 10021, USA

### ARTICLE INFO

#### Article history:

Received 2 November 2015  
 Received in revised form 11 July 2016  
 Accepted 12 July 2016  
 Available online 14 July 2016

#### Keywords:

Nanoparticle  
 PRINT  
 cGMP delivery vehicle  
 Liquidia technologies  
 Team-based science  
 Clinical trials  
 Particle engineering  
 Drug delivery vehicle  
 Particulate vaccine  
 Calibration quality particles

### ABSTRACT

Nanoparticle properties such as size, shape, deformability, and surface chemistry all play a role in nanomedicine drug delivery. While many studies address the behavior of particle systems in a biological setting, revealing how these properties work together presents unique challenges on the nanoscale. Particle replication in non-wetting templates (PRINT®) is one molding technique that allows for fabrication of “calibration quality” micro and nanoparticles with independent control over their physical parameters. As the only technology in the world capable of independently optimizing and robustly manufacturing GMP compliant precision particles of virtually any size, shape, and composition, the PRINT technology has the capability to engineer the future of healthcare.

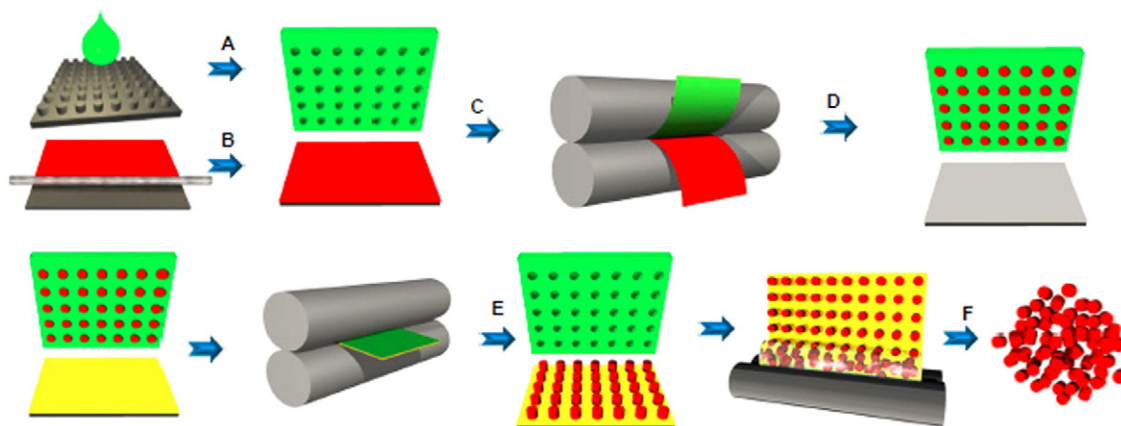
© 2016 Published by Elsevier B.V.

Polymer Science is at the “Core” of all we do. When my group began reporting on the synthesis of novel, functionalized, photocurable perfluoropolyethers (PFPEs) in supercritical carbon dioxide in the late 1990s, we called these materials “Liquid Teflon” [1,2]. These functional PFPE materials could be photochemically crosslinked at room temperature to yield clear, flexible, organic-solvent resistant materials that could be utilized for a variety of applications including for use in high-resolution imprint lithography. By employing these novel materials and adapting manufacturing approaches from the microelectronics industry, my group invented Particle Replication in Non-wetting Templates (PRINT®), a nano-materials manufacturing process that, for the first time, enabled the scalable fabrication of large batches of highly uniform, shape-specific nano- and microparticles from almost any chemical composition (Fig. 1) [3–10]. Three important advantages of PFPEs enhance the performance of PRINT-embossed molds: i) the low surface energy of PFPEs allow using any organic liquid to selectively fill only the mold's cavities—without wetting the land area between the cavities—eliminating the problematic interconnecting “flash layer”; ii) organic liquids that are “pre-particle” solutions do not swell fluoropolymers; and iii) Teflon-like characteristics of PFPE molds

allow for the easy removal (harvesting) of the particles from mold cavities. Thus, PRINT allows for the fabrication of particles with precise control over size (50 nm to >100 μm), shape, modulus, composition (hydrogels, thermoplastic polymers, active therapeutics, proteins, nucleic acids, imaging agents), and surface chemistries (antibodies, “stealth” agents).

As the numerous applications for PRINT began to unfold, my group expanded to integrate expertise from multiple scientific fields (e.g. chemistry, engineering, biology, immunology, pharmaceutical sciences, medicine). As my talented, diversified team optimized this technology, important advances were made in understanding the interactions of particles in a biological setting. This knowledge provided direction to develop PRINT-based chemotherapeutic nanoparticles for treating cancer [11–16], a PRINT-based microneedle drug delivery system for a variety of therapeutic applications [17], and a PRINT nanoparticle-based vaccine platform [18]. PRINT has not only provided valuable insight into particle parameters essential for designing successful therapeutics, but through its development has also underscored a successful way to achieve innovation in the lab. The development and evolution of PRINT brought together talented, knowledgeable, and hard-working individuals with a wide range of academic backgrounds and life experiences—and stemming from these backgrounds and experiences—unique interests, perspectives, skills, capabilities, and problem-solving approaches. It is

E-mail address: [desimone@email.unc.edu](mailto:desimone@email.unc.edu).



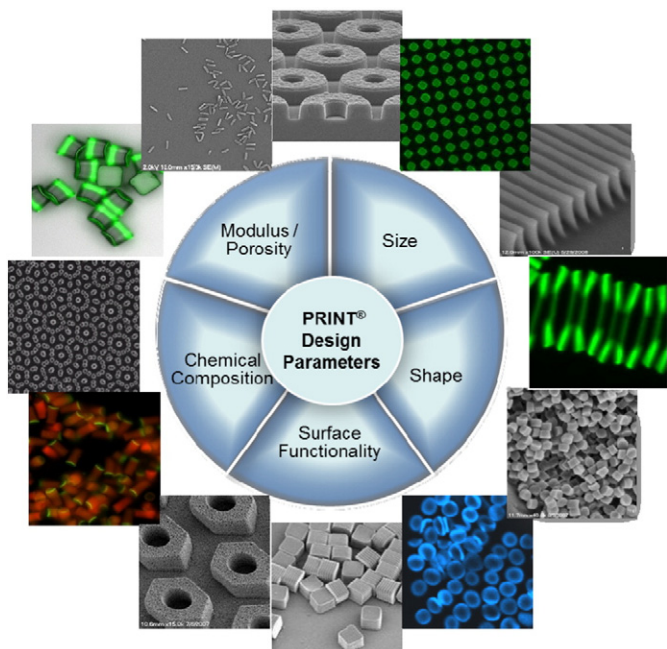
**Fig. 1.** The PRINT Process: (A) A liquid polymer mold (green) is delivered onto a silicon master with the desired size and shape of the particle (grey), the polymer is then photo-chemically cross-linked and peeled away to generate a mold having cavities; (B) solution containing desired particle components (red) is distributed on a delivery sheet and then is (C) transferred to the mold (green) by a roller; (D) material in mold cavities is solidified in the mold; (E) particles are removed from the mold with a harvesting film (yellow); (F) the particles are then freed from the harvest film to provide free-flowing particles.

my opinion that there is no more fertile ground for innovation than a diversity of experience. A successful scientific endeavor is one that attracts a diversity of experience, draws upon the breadth and depth of that experience, and cultivates those differences, acknowledging the creativity they spark.

The PRINT technology evolved into a robust and reproducible platform to rationally design precise particles with virtually any size, shape, surface functionality, mechanical modulus and chemical composition to target therapeutic safety and efficacy (Fig. 2). Depending on the therapeutic application, PRINT particles can be tailored in size from tens of nanometers to hundreds of micrometers, in a variety of particle geometries, and with high degree of monodispersity. For example, PRINT particles smaller than 100 nm (in at least one dimension) are the “top performers” for intravenous applications while 1  $\mu\text{m}$  particles have been identified as optimal for inhaled administration in pulmonary applications. The ability to manufacture PRINT particles of sizes 10 nm or smaller are limited due to the capabilities of current state of the art photolithographic techniques. Advances in lithography will be needed to

generate smaller feature size master templates that can then be utilized in the PRINT process. The chemical flexibility of the PRINT technology offers many potential advantages in formulating both small molecule active pharmaceutical ingredients (API) and biologic API drug delivery systems such as targeting chemistries, improved drug/cargo loading, and delivery of sensitive cargos like peptides and nucleic acid, and combination products. In each case, the goal is to engineer a particle design that offers a novel, safe and cost-effective product and offers strong new product intellectual property. As the only technology in the world capable of independently designing and manufacturing precision particles of any size, shape and chemistry, the PRINT platform has the agility required to meet the product development needs of the healthcare industry and consistency required to address increasing regulatory demands.

Liquidia Technologies, the company founded based on PRINT, has evolved the technology to be “cGMP compliant,” required for use in U.S. human clinical trials. In order to realize the potential of the PRINT technology, scale needed to be addressed. As shown in Fig. 1, the particle fabrication process consists of a series of steps: 1) coating material, 2) filling material, and 3) collecting particles. It was realized that these types of steps were readily adaptable to a roll-to-roll process. This idea translated into a custom built roll-to-roll machine for the production of PRINT particles as shown in Fig. 3. Liquidia Technologies has invested significant effort and resources in the scale up of the PRINT manufacturing platform since the early stages of the company. Specifically during 2010–2013, with support from a cost-sharing grant awarded through the National Institute for Standards and Technology (NIST), Liquidia set a goal to demonstrate a 1000-fold throughput improvement for fabrication of nanoparticles. This was accomplished by increasing the



**Fig. 2.** PRINT technology design parameters: size, shape, surface functionality, chemical composition and modulus/porosity.



**Fig. 3.** Custom-built equipment for roll-to-roll production of nanoparticles.

width of particle fabrication, the speed of the process, and the efficiency of the process. These approaches were undertaken using a combination of experimental prototyping, testing and optimization in combination with extensive computational modeling of the system. Completion of this effort in January 2013 was accompanied by the successful demonstration of 1000-fold increase in nanoparticle throughput, but perhaps more importantly, the full integration of multiple processing improvements into routine use at Liquidia, resulting in a dramatic increase in manufacturing capacity. These improved capabilities have provided Liquidia with sufficient capacity to support multiple pre-clinical and clinical development efforts. This production and development line has been used for the production of vaccine nanoparticles at cGMP compliant standards in the manufacture of LIQ001, a novel, and first generation influenza vaccine candidate evaluated in human clinical trials in 2010. The initial phase I/IIa clinical trial in influenza demonstrated the cGMP compliance and safety of PRINT particles [19]. Additionally, this platform can be scaled to satisfy potential global manufacturing needs.

Due to the ubiquitous nature of nanoparticles in the field of drug delivery, the PRINT technology has been utilized across not only different therapeutic areas, but also across multiple companies focusing on various diseases. Liquidia Technologies pioneered the development of the PRINT technology focused on inhalation and vaccine therapeutic areas. These advancements led to a partnership developing products in collaboration with GlaxoSmithKline [20]. In 2013, Envisia Therapeutics, an ophthalmology focused company, spun out of Liquidia Technologies. Using the PRINT technology, Envisia is rationally designing PRINT particle therapeutics that are precisely engineered to overcome the biologic, chemical or other barriers that limit existing ocular therapies [21]. In addition, in 2014 Liquidia spun out Lq3 Pharmaceuticals, which is leveraging the unique benefits of the PRINT technology to develop innovative products in oral health [22].

Our research continues to operate at the interface of polymer chemistry, nanotechnology, and medicine with the purpose of engineering better therapies, vaccines, and medical devices. The advent of “calibration quality” particles using PRINT has provided a means for investigating and understanding the interdependent role of particle size, shape, surface functionality, and matrix composition on the fate of organic nanoparticles in biological settings. We continue to pursue therapeutic applications and, subsequently, utilize the agility of PRINT to elucidate particle parameters that are crucial for enhancing intended particle biodistribution, specific cellular interactions, manipulating intracellular location of particles, and minimizing cytotoxic “bystander” effects based upon target site and route of administration. As the only technology in the world capable of independently optimizing and robustly manufacturing precision particles of virtually any size, shape, and composition, the PRINT technology has the capability to engineer the future of healthcare.

## Acknowledgements

I would like to thank Liquidia Technologies (Ashley Galloway), Envisia Therapeutics (Stuart Williams, Andres Garcia), Crista Farrell and Chris Luft of UNC-CH for the assistance in composing this review. This work was supported by numerous funding agencies and collaborations including

Liquidia Technologies, the Carolina Center for Cancer Nanotechnology Excellence U54CA151652, NIH Pioneer Award 1DPOD006432, NIH Grant U19AI109784, and Defense Threat Reduction Agency (DTRA) Award HDTRA1-13-1-0045.

## References

- [1] W.C. Bunyard, T.J. Romack, J.M. DeSimone, Perfluoropolyether synthesis in liquid carbon dioxide by Hexafluoropropylene Photooxidation, *Macromolecules* 32 (1999) 8224–8226.
- [2] J.P. Rolland, R.M. Van Dam, D.A. Schorzman, S.R. Quake, J.M. DeSimone, Solvent resistant photocurable “Liquid Teflon” for microfluidic device fabrication, *J. Am. Chem. Soc.* 126 (8) (2004) 2322–2323.
- [3] J.P. Rolland, B.W. Maynor, L.E. Euliss, A.E. Exner, G.M. Denison, J.M. DeSimone, Direct fabrication and harvesting of monodisperse, shape specific nano-biomaterials, *J. Am. Chem. Soc.* 127 (2005) 10096–10100.
- [4] R.A. Petros, P.A. Ropp, J.M. DeSimone, Reductively labile PRINT particles for the delivery of doxorubicin to HeLa cells, *J. Am. Chem. Soc.* 130 (2008) 5008–5009.
- [5] J.Y. Kelly, J.M. DeSimone, Shape-specific, monodisperse nano-molding of protein particles, *J. Am. Chem. Soc.* 130 (2008) 5438–5439.
- [6] L. Euliss, J. DuPont, J.M. DeSimone, Imparting size, shape, and composition control of materials for nanomedicine, *Chem. Soc. Rev.* 35 (2006) 1095–1104.
- [7] S.E.A. Gratton, S.S. Williams, M.E. Napier, P.D. Pohlhaus, Z. Zhou, K.B. Wiles, B.B. Maynor, C. Shen, T. Olafsen, E.T. Samulski, J.M. DeSimone, The pursuit of a scalable nano-fabrication platform for use in material and life science applications, *Acc. Chem. Res.* 41 (2008) 1685–1695.
- [8] J.L. Perry, K.P. Herlihy, M.E. Napier, J.M. DeSimone, PRINT: a novel platform toward shape and size specific nanoparticle therapeutics, *Acc. Chem. Res.* 44 (10) (2011) 990–998.
- [9] S.E.A. Gratton, P.A. Ropp, P.D. Pohlhaus, J.C. Luft, V.J. Madden, M.E. Napier, J.M. DeSimone, The effect of particle design on cellular internalization pathways, *Proc. Natl. Acad. Sci.* 105 (33) (2008) 11613–11618.
- [10] R.A. Petros, J.M. DeSimone, Strategies in the design of nanoparticles for therapeutic applications, *Nat. Rev. Drug Discov.* 9 (2010) 615–627.
- [11] J. Xu, R.A. Petros, M.E. Napier, J.M. DeSimone, The complex role of multivalency in nanoparticles targeting the transferrin receptor for cancer therapies, *J. Am. Chem. Soc.* 132 (32) (2010) 11306–11313.
- [12] S. Dunn, S. Tian, S. Blake, J. Wang, A. Galloway, A. Murphy, P. Pohlhaus, J. Rolland, M. Napier, J.M. DeSimone, Reductively-responsive siRNA-conjugated hydrogel nanoparticles for Gene silencing, *J. Am. Chem. Soc.* 134 (2012) 7423–7430.
- [13] M. Parrott, M. Finniss, J. Luft, A. Pandya, A. Gullapalli, M. Napier, J.M. DeSimone, Incorporation and controlled release of Silyl ether pro-drugs from PRINT nanoparticles, *J. Am. Chem. Soc.* 134 (2012) 7978–7982.
- [14] J. Xu, J. Wang, J. Luft, S. Tian, G. Owens, A. Pandya, P. Berglund, P. Pohlhaus, B. Maynor, M. Napier, J.M. DeSimone, Rendering protein-based particles transiently insoluble for therapeutic applications, *J. Am. Chem. Soc.* 134 (2012) 8774–.
- [15] K.S. Chu, A.N. Schorzman, M.C. Finniss, C.J. Bowerman, L. Peng, J.C. Luft, A. Madden, A.Z. Wang, W.C. Zamboni, J.M. DeSimone, Nanoparticle drug loading as a design parameter to improve Docetaxel pharmacokinetics and efficacy, *Biomaterials* 34 (33) (2013) 8424–8429.
- [16] K.S. Chu, M.C. Finniss, A.N. Schorzman, J.L. Kuijter, J.C. Luft, J. Bowerman, M.E. Napier, Z.A. Haroon, W.C. Zamboni, J.M. DeSimone, PRINT nanoparticles with tumor selective alkyl silyl ether Docetaxel Prodrug reduces toxicity, *Nano Lett.* 14 (3) (2014) 1472–1476.
- [17] K.A. Moga, L.R. Bickford, R.D. Geil, S.S. Dunn, A.A. Pandya, Y. Wang, J.H. Fain, C.F. Archuleta, A.T.O. O’Neill, J.M. DeSimone, Rapidly-dissolvable microneedle patches via a highly scalable and reproducible soft lithography approach, *Adv. Mater.* 25 (36) (2013) 5060.
- [18] C.A. Fromen, G.R. Robbins, T.W. Shen, M.P. Kai, J.P.Y. Ting, J.M. DeSimone, Controlled analysis of nanoparticle charge on mucosal and systemic antibody responses following pulmonary immunization, *Proc. Natl. Acad. Sci.* 112 (2) (2015) 488–493.
- [19] A.L. Galloway, A. Murphy, J.M. DeSimone, J. Di, J.P. Hermann, M.E. Hunter, J.P. Kindig, F.J. Malinoski, M.A. Rumley, D.A. Stoltz, T.S. Templeman, B. Hubby, Development of a nanoparticle-based influenza vaccine using the PRINT technology, *Nanomed. Nanotechnol. Biol. Med.* 9 (2013) 523–531.
- [20] www.liquidia.com
- [21] www.envisiatherapeutics.com
- [22] www.lq3pharma.com